



## Production of protease enzyme by *Enterobacter sakazakii* using solid state fermentation

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### ABSTRACT

With a full-factorial facing centered central composite design, response surface approach was used. To enhance protease production, two arrangements of tests were completed to upgrade the five most significant components of the development conditions. In carafes, the hatching time, temperature, introductory media pH, and level of rice wheat supplement to the media were completely tuned, while in a bioreactor, the fomentation and air circulation impact were controlled. This plan diminished the quantity of genuine trials expected to streamline protease blend and took into account the examination of likely connections between boundaries. The immediate and quadratic effects of starting still uncertain in the principle plan of fundamentals Protease creation is impacted by the primary part. Both straight effects of disrupting and air dissemination in the bioreactor were considered to be significantly colossal (> near 100%) to the extent protease yield. At a temperature of 39.7 oC, an essential pH of 7.1, supplementation with 13.6 percent rice wheat, upsetting at 320 rpm, and wind current at 0 vvm, ideal protease blend was found.

### Keywords:

Bacterial potease ,enterobacter sakazakii cultivation conditions

### Introduction

*Cronobacter sakazakii*, recently known as *Enterobacter sakazakii*, is a Gram-negative, pole formed, pathogenic microscopic organisms that can get by in amazingly dry conditions <sup>(1)</sup>. The majority of *Cronobacter sakazakii* cases occur in adults, but the most vulnerable are low-birth-weight 'preterm newborn babies and older infants <sup>(2)</sup>. More than 120 cases of *C. sakazakii*-related disease have been reported, with the majority of them being life-threatening (FAO/WHO, 2008). Many of these outbreaks have been linked to the ingestion of *C. sakazakii*-infected powdered infant formula, resulting in a slew of recalls and legal action. Proteases account for 60–65 percent of the global industrial market <sup>(3)</sup> Numerous *Cronobacter* species produce a scope of extracellular proteases, which are fundamental for cell work. Protease is a reactant chemical that separates

proteins into more modest pieces or single amino acids, permitting new protein items to create <sup>(4)</sup> Proteases are a single class of enzymes that are widely employed in detergents, medicines, leather, food, and agriculture. <sup>(5)</sup>

For the generation of microbial metabolites, solid-state fermentation is a typical method <sup>(6)</sup> The SSF interaction is done on a strong substrate with a low dampness content, bringing about a high item focus with simply a modest quantity of energy required <sup>(7)</sup>.

### Materials and Methods

#### Chemicals

Hi-Media Laboratories, Merck (Mumbai, India), and Sigma provided all of the compounds utilized in this investigation (all of which were 99 percent pure) (U.S.A).

### Culture and Growth Condition

*C. sakazakii* was gotten from the Biotechnology Department of AL-Nahrain University's College of Science in Iraq. The strain was sub-refined at regular intervals on healthful agar inclines with a pH of 7.0 at 37C inclinations.

### Production of enzyme

Subsequent to streamlining the upsides of different wholesome and actual variables in confinement, the five most significant elements as far as *Enterobacter sakazakii* protease creation, in particular rice wheat fixation (X1), hatching temperature (X2), starting pH (X3), unsettling speed (X4), and air circulation volume (X5), were recognized.

### Screening ability of *Cronobacter sakazakii* isolates for protease production

#### Semi quantitative screening

*C. sakazakii* secludes were streaked on wholesome agar medium and refined at 37C for 24 hours. From that point onward, one state was chosen and put on a skim milk agar medium plate. The plate was brooded for 24 hours at 37C. The presence of an unmistakable corona zone around every province was utilized to survey *C. sakazakii*'s capacity to deliver proteases.

#### Quantitative screening of protease

Following the methods below, production was carried out by determining the enzymatic activity and specific activity.

#### Assay of protease enzyme Activity

Protease development was directed by incubating 0.8 ml of 1% casein course of action with 2.0 ml of enzymatic response for 30 minutes at 37 0C, as depicted by Manachini et al. (1989). The reaction was finished by adding 1 mL of 5% TCA and cooling the course of activity in an ice shower for 10-15 minutes prior to centrifuging it at 6000 rpm for 15 minutes. The control test was made by blending 1 ml of 5% TCA with 0.8 ml of 1% casein strategy, then, at that point, adding 0.2 ml enzymatic game-plan. An UV-VIS spectrophotometer was utilized to test the absorbance of the supernatant at 280 nm. The breakdown of casein protein to short peptides and dissolvable amino acids was utilized to overview enzymatic movement, since one unit (U) of compound action was portrayed as the proportion of driving force fundamental

to incite an augmentation in absorbance at 280 nm indistinguishable from 0.01 in one second under exploratory conditions, the absorbance was evaluated at 280 nm.

#### Compound movement :Absorbance at 280nm%0.01\*30\*0.2

0.01: Constant

30: Reaction time (min)

0.2: Enzyme volume (ml)

#### Unequivocal development (unit\mg):

#### Enzyme activity%Protien center

#### Protein evaluation

The measure of protein in each example was determined utilizing the accompanying equation (Bradford, 1976):

#### Optimization of media

Distinctive actual elements including as pH, temperature, inoculum size, brooding length, and fomentation were advanced in the media utilized for protease amalgamation.

#### Ideal inoculum size

The impact of different inoculum sizes of the picked over maker disengage on protease creation was examined. This was refined by exclusively immunizing the creation medium with an inoculum size going from (1102-1108) CFU/ml.

#### Ideal pH

The pH of the creation medium was ideally acclimated to various pH esteems to look at the impact of medium pH on protease amalgamation by the disconnect *C.sakazakii* (4, , 5, , 6, , 7, , 8, and 9). In the accompanying period of enhancement, the ideal pH was utilized to recharge the creation medium.

#### Ideal brooding temperature

The creation medium was brooded at a few hatching temperatures (25, 30, 35, 40, 45, and 50C) to build up the best temperature for protease combination by disengage of *C.sakazakii*, and afterward the ideal hatching temperature was set at the accompanying periods of advancement.

#### Ideal brooding period

The impact of hatching time on *C.sakazakii* separate protease creation was researched by brooding the creation mode for different timeframes (12, 24, 36, 48, and 60 hours) to track down the ideal time frame for catalyst

combination. The ideal brooding span still up in the air in the resulting phases of advancement.

**Ideal disturbance speed**

The ideal shaking speed for protease blend by the picked tie was coordinated by anguishing the creation medium at different shaking speeds (100, 140, 150, 200, and 250 rpm).

**Results**

The sythesis of rice grain, hatching temperature, beginning pH, fomentation, and air circulation were displayed to greatestly affect the amalgamation of protease proteins by *Enterobacter sakazakii* ASUIA279 in the starter

examination. The tests were parted into two sets because of gear and time limitations, with the initial three parts upgraded in an Erlenmeyer carafe. In a 2 L bioreactor, disturbance and air circulation were tuned utilizing the ideal culture boundaries set up from the principal set of preliminaries. Following 5 days of aging, the maximal protease debasing catalyst action was found, and the reaction was gathered by then. Tables 2 and 3 portray the discoveries for the two stages, depict the protease-debasing catalyst action (reaction) of the examinations

Variables	Symbol coded	Range and level		
		-1	0	+1
Temperature (°C)	X1	25	35	50
Initial pH	X2	4	6	9
Rice bran % (w/v)	X3	4	7	12
Agitation (rpm)	X4	100	150	250
Aeration (vvm)	X5	0	1	2

**Table 1.** Test reach and levels of the free factors for protease-corrupting catalyst Production

Experiment run no.	Temperature (°C)	pH	% Rice Bran	Experimental Protease Activity (U/ml)
1	25	4	5	11.006
2	30	5	10	6.475
3	35	6	10	7.336
4	40	7	5	11.530
5	45	8	15	4.300
6	50	9	15	12.654

**Table 2.** Experimental design of three independent variables in step-one showing experimental responses

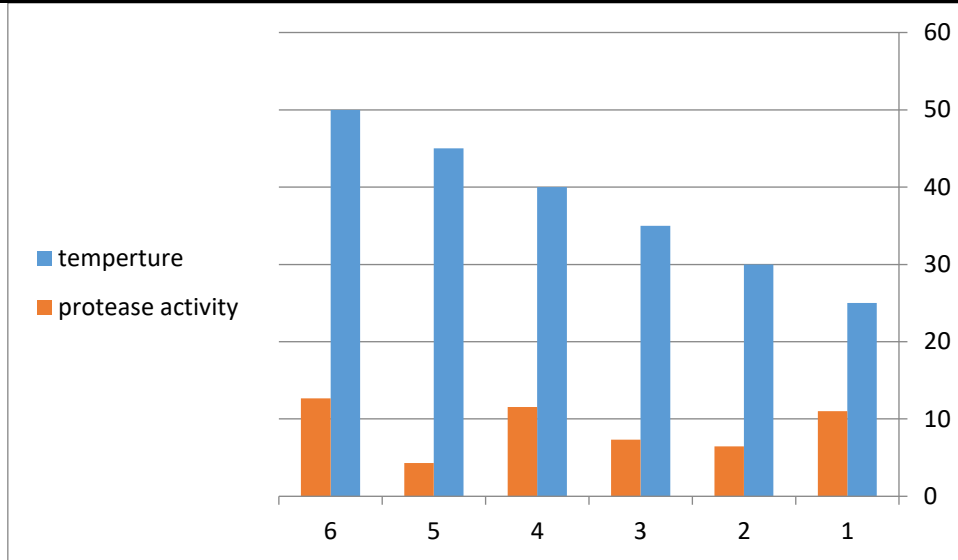


Table 1: relationship between temperature and protease activity show high activity in 50 c

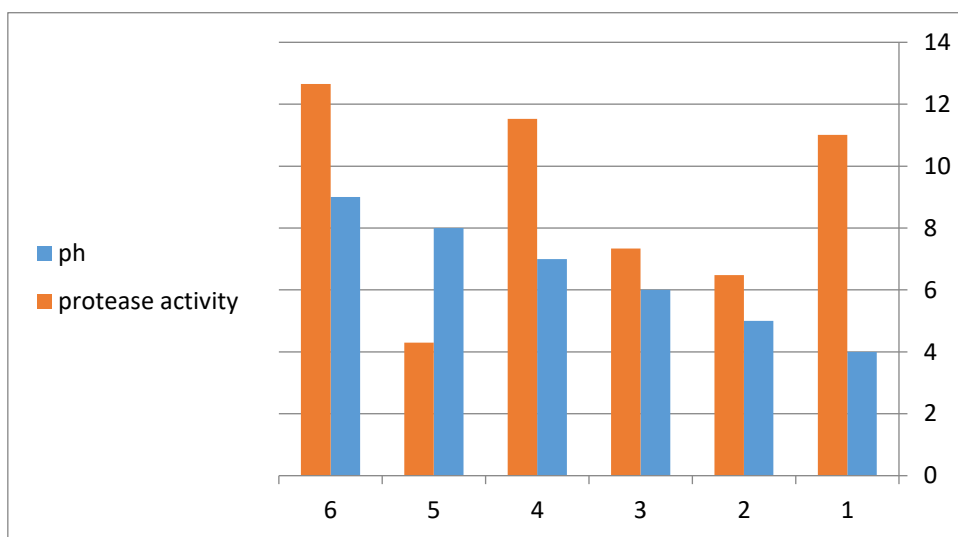


Table 2: relationship between pH and protease activity show high activity in pH 12

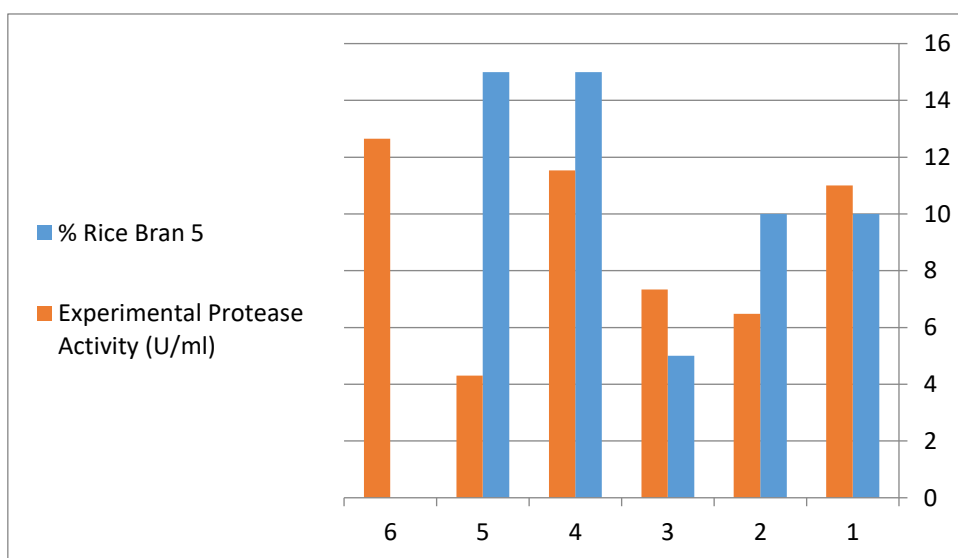


Table 3 : relationship between rice bran and protease activity show high activity in 50 c

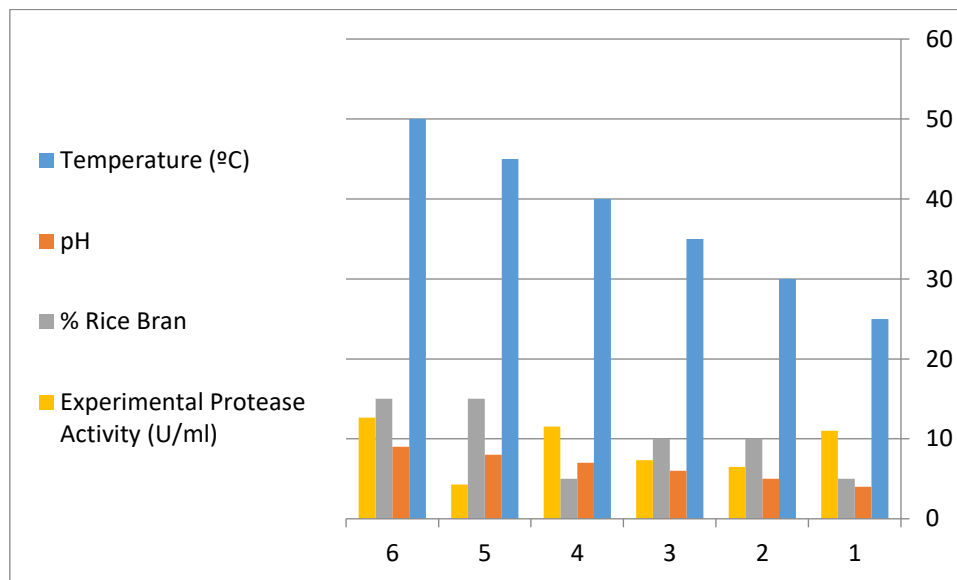


Table 4: relationship between temperature ,pH , rice bran and protease activity .

## Discussion

Because of the associations among the components, conventional methodologies for multifaceted trial configuration are tedious and unequipped for accomplishing the authentic ideal <sup>(9)</sup> . The source and ideal grouping of carbon are significant elements in the creation of protease-debasing catalyst, and pH, temperature, unsettling, broke down oxygen, and strain are the main actual boundaries that fundamentally affect the development of creatures and the creation of metabolites from them <sup>(10)</sup> .

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